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## THE VIRUS RANGE OF HOP PLANTS: IDENTIFICATION KEY

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The term "virus range" was coined by Christie and Crawford in 1978, by analogy with the term "host range", to denote all viruses that can infect a particular host species [1]. Studying the virus range of a particular host species, similar to investigating the host range of a particular virus, plays an essential role in improving the understanding of virus-host-environment interactions [2]. However, investigating the virus range of a particular host necessitates employing approaches that do not rely on *a priori* information about which viruses might be present in the sample to detect all viruses it contains. One of these approaches involves the preliminary inspection of samples under the transmission electron microscope (TEM), followed by identifying the observed viruses using serological and nucleic acid-based techniques [3]. However, identifying the virus species observed in a sample under the TEM also requires the development of identification keys, based on the shape and size of virions.

The viral diseases of the common hop (*Humulus lupulus* L.) have been studied for centuries. Numerous authors in different periods have reported various numbers of viral diseases of hop and virus species infecting this plant. For example, Schmidt and Klinkowski listed 11 viroses of hop in Europe in 1965 [4]. However, before the implementation of serological techniques, the causative agents of hop viroses were characterized by diagnostic species and a combination of symptoms of the disease they cause in those species and the hop themselves. Hence, determining the taxonomic position of the pathogens that caused diseases described during that period, such as those documented by Schmidt and Klinkowski, is impossible. For example, Adams and others revealed that arabis mosaic virus (ArMV) causes hop chlorotic disease in England, whereas Eppler indicates hop mosaic virus (HMV) as the causative agent of this disease in Germany [5, 6]. Nevertheless, many authors identified numerous virus species that infect the common hop and can be re-identified by other researchers by using serological and nucleic acid-based techniques. Thus, Pethybridge and colleagues performed a meta-analysis of studies on viruses infecting hop plants and listed 15 identifiable virus species [7].

Therefore, the study aimed to gather information about the virion size and shape of viruses belonging to the virus range of the common hop and to develop the identification key. To this end, the author was searching for information about the virion size and shape of viruses listed by Pethybridge and others [7]. The compiled information is presented in the table.

**Table 1.** The virion size and shape of viruses infecting hop

| Species                                 | Shape                  | Size, nm           | Reference |
|---|------------------------|--------------------|-----------|
| Alfalfa mosaic virus, AIMV              | bacilliform            | 30 - 56 × 19*      | 7         |
|   | bacilliform            | 61 × 17.8**        | 8         |
| American hop latent virus, AHLV         | filamentous            | 676 × 14.4         | 9         |
| Apple mosaic virus, ApMV                | quasi-isometric        | 24 × 26 - 28       | 10        |
|   | isometric              | 24                 | 10        |
|   | isometric              | 22                 | 10        |
|   | isometric              | 25                 | 11        |
|   | isometric              | 32                 | 11        |
|   | quasi-isometric        | 30 × 30 - 36       | 11        |
| Arabidopsis mosaic virus, ArMV          | isometric              | 25 - 30*           | 7         |
|   | polyhedral             | 30***              | 12        |
| Cherry leaf roll virus, CLRV            | isometric              | 25 - 30*           | 7         |
| Cucumber mosaic virus, CMV              | isometric              | 25-30*             | 7         |
|   | isometric              | 30***              | 12        |
| Hop latent virus, HLV                   | filamentous            | 678 × 15.2         | 9         |
|   | filamentous            | 650 × 13           | 10        |
| Hop mosaic virus, HMV                   | filamentous            | 651 × 13.8         | 13        |
|   | filamentous            | 625 × 13           | 14        |
|   | filamentous            | 655 × 13.5         | 8         |
|   | filamentous            | 630 - 680 × 2<br>8 | 15        |
| Humulus japonicus latent virus, HJLV    | quasi-isometric        | 24 - 33            | 16, 17    |
| Petunia asteroid mosaic virus, PeAMV    | isometric              | 32 - 35*           | 7, 18, 19 |
| Prunus necrotic ringspot virus, PNRSV   | isometric, bacilliform | 23, 25, 27*        | 7         |
|   | isometric              | 40                 | 20        |
|   | isometric              | 28 - 30            | 21        |
|   | quasi-isometric        | 25**               | 22        |
|   | quasi-isometric        | 31**               | 22        |
| Raspberry bushy dwarf virus, RBDV       | isometric              | 33*                | 7         |
| Strawberry latent ringspot virus, SLRSV | isometric              | 30*                | 7         |
| Tobacco mosaic virus, TMV               | rod-shaped             | 300 × 18*          | 7         |
| Tobacco necrosis virus, TNV             | isometric              | 28*                | 7         |
|   | isometric              | 26                 | 23        |
|   | isometric              | 27 - 28            | 21        |
| Tobacco ringspot virus, TRSV            | isometric              | 25 - 30*           | 7         |

Notes: \*The size and shape are listed for the virus species in general, not for the strain infecting hop [7]; \*\*The sizes of virions were measured by the author using the website <https://eleif.net/photomeasure> and the micrograph published in the paper by Yu and Liu, as the authors did not specify the virion sizes [8]. The size is indicated as the average value for 5 virions and was calculated in relation to the size of the bar; \*\*\*The author consistently confuses  $\mu\text{m}$  with nm in its publication.

According to the table, the viruses infecting hop plants can be distinguished by reference to the following identification key.

**Identification key to the virus range of hop plants**

1(2). Virions are isometric and / or quasi-isometric and / or bacilliform, with the biggest dimension being less than 100 nm. – Group of isometric, quasi-isometric, and bacilliform viruses (AIMV, ApMV, ArMV, CLRV, CMV, HJLV, PeAMV, PNRSV, RBDV, SLRSV, TNV and TRSV).

2(3). Virions are rod-shaped, ca. 300 nm in length. – TMV.

3. Virions are filamentous, longer than 500 nm. – *Carlavirus complex* (AHLV, HLV, and HMV).

The identification results obtained from the key should be confirmed or specified by serological or nucleic acid-based techniques. In future work, the identification key can be supplemented with new virus species infecting hop plants if they are discovered.

**Reference:**

1. Christie S. R., Crawford W. E. Plant virus range of *Nicotiana benthamiana*. Plant Disease Reporter. 1978. Vol. 62. № 1. P. 20-22. <https://books.google.com/books?id=6gcN7xqcNwUC&pg=PA20>
2. Roossinck M. J. Plants, viruses and the environment: Ecology and mutualism. Virology. 2015. Vol. 479-480. P. 271-277. <http://dx.doi.org/10.1016/j.virol.2015.03.041>
3. Whattam M., Dinsdale A., Elliott C. E. Evolution of Plant Virus Diagnostics Used in Australian Post Entry Quarantine. Plants. 2021. Vol. 10. №7. P. 1430. <http://dx.doi.org/10.3390/plants10071430>
4. Schmidt H. E., Klinkowski M. Virosen des Hopfens (*Humulus lupulus* L.) in Europa. Journal of Phytopathology. 1965. Vol. 54. №2. P. 122-146. <http://dx.doi.org/10.1111/j.1439-0434.1965.tb04087.x>
5. ADAMS A. N., BARBARA D. J., VIES D. L. D. The etiology of hop chlorotic disease. Annals of Applied Biology. 1987. Vol. 111. №2. P. 365-371. <http://dx.doi.org/10.1111/j.1744-7348.1987.tb01464.x>
6. Eppler A. ARABIS MOSAIC VIRUS IN GERMAN HOPS. Acta Horticulturae. 1992. №308. P. 81-86. <http://dx.doi.org/10.17660/actahortic.1992.308.9>
7. Pethybridge S. J., Hay F. S., Barbara D. J., Eastwell K. C., Wilson C. R. Viruses and Viroids Infecting Hop: Significance, Epidemiology, and Management. Plant Disease. 2008. Vol. 92. №3. P. 324-338. <http://dx.doi.org/10.1094/pdis-92-3-0324>
8. Sastry K. S., Mandal B., Hammond J., Scott S. W., Briddon R. W. Encyclopedia of Plant Viruses and Viroids. New Delhi: Springer India, 2019. 2936 p. <https://doi.org/10.1007/978-81-322-3912-3>
9. YU J., LIU Y. The occurrence of three viruses in hop (*Humulus lupulus*) in China. Plant Pathology. 1987. Vol. 36. №1. P. 38-44. <http://dx.doi.org/10.1111/j.1365-3059.1987.tb02175.x>
10. ADAMS A. N., BARBARA D. J. Host range, purification and some properties of two carlaviruses from hop (*Humulus lupulus*): hop latent and American hop latent. Annals of Applied Biology. 1982. Vol. 101. №3. P. 483-494. <http://dx.doi.org/10.1111/j.1744-7348.1982.tb00849.x>
11. KANNO Y., YOSHIKAWA N., TAKAHASHI T. Some Properties of Hop Latent and Apple Mosaic Viruses Isolated from Hop Plants and Their Distributions in Japan. Japanese Journal of Phytopathology. 1993. Vol. 59. №6. P. 651-658. <http://dx.doi.org/10.3186/jjphytopath.59.651>
12. SANO T., SASAKI M., SHIKATA E. Apple mosaic virus isolated from hop plants in Japan. Annals of Applied Biology. 1985. Vol. 106. №2. P. 305-312. <http://dx.doi.org/10.1111/j.1744-7348.1985.tb03120.x>

13. BOCK K. R. Arabis mosaic and Prunus necrotic ringspot viruses in hop (*Humulus lupulus* L.). *Annals of Applied Biology*. 1966. Vol. 57. №1. P. 131-140. <http://dx.doi.org/10.1111/j.1744-7348.1966.tb06873.x>
14. ADAMS A. N., BARBARA D. J. Host range, purification and some properties of hop mosaic virus. *Annals of Applied Biology*. 1980. Vol. 96. №2. P. 201-208. <http://dx.doi.org/10.1111/j.1744-7348.1980.tb02980.x>
15. KANNO Y., IIDA H., YOSHIKAWA N., TAKAHASHI T. Some Properties of Hop Mosaic Virus Isolated in Japan. *Japanese Journal of Phytopathology*. 1994. Vol. 60. №6. P. 675-680. <http://dx.doi.org/10.3186/jjphytopath.60.675>
16. Macovei A. Vorkommen mechanisch übertragbarer Viren an Hopfen (*Humulus lupulus*L.) in der Sozialistischen Republik Rumänien. *Archives Of Phytopathology And Plant Protection*. 1976. Vol. 12. №2. P. 75-86. <http://dx.doi.org/10.1080/03235407609431735>
17. ADAMS A. N., CLARK M. F., BARBARA D. J. Host range, purification and some properties of a new ilarvirus from *Humulus japonicus*. *Annals of Applied Biology*. 1989. Vol. 114. №3. P. 497-508. <http://dx.doi.org/10.1111/j.1744-7348.1989.tb03365.x>
18. Scott S. W., Zimmerman M. T. The complete sequence of the genome of *Humulus japonicus* latent virus. *Archives of Virology*. 2006. Vol. 151. №8. P. 1683-1687. <http://dx.doi.org/10.1007/s00705-006-0771-6>
19. Novák J. B., Lanzová J. Identification of alfalfa mosaic virus and tomato bushy stunt virus in hop (*Humulus lupulus* L.) and grapevine (*Vitis vinifera* subsp.sativa (DC./HEGI) plants in czechoslovakia. *Biologia Plantarum*. 1976. Vol. 18. №2. P. 152-154. <http://dx.doi.org/10.1007/bf02923156>
20. Lovisollo O., Bode O., Völk J. Biologische Bundesanstalt für Land- und Forstwirtschaft, Institut für landwirtschaftliche Virusforschung, Braunschweig. *Journal of Phytopathology*. 1965. Vol. 53. №4. P. 323-342. <http://dx.doi.org/10.1111/j.1439-0434.1965.tb04081.x>
21. Jochimsen K. N. Evaluation of hop (*Humulus lupulus* L.) protoplast inoculation with *Prunus necrotic ringspot virus* : Masters Thesis. Corvallis, 1987. 130 p. <http://hdl.handle.net/1957/40201>
22. Macovei A. A survey of the Romanian hop viruses and their characterisation by electron microscopy. *Proceedings of the International Workshop on Hop Virus Diseases, Rauischholzhausen Castle, 27.06.-02.07.1988* / ed. by A. Eppler, Deutsche Phytomedizinische Gesellschaft. Hohenheim, 1989. P. 19-22. <https://books.google.com/books?id=JBNIAAAAYAAJ>
23. BOCK K. R. Strains of *Prunus necrotic ringspot virus* in hop (*Humulus lupulus* L.). *Annals of Applied Biology*. 1967. Vol. 59. №3. P. 437-446. <http://dx.doi.org/10.1111/j.1744-7348.1967.tb04460.x>
24. Chod J., Jokeš M., Novák M. The electron-microscopic proof of tobacco necrosis virus in hop plant. *Biologia Plantarum*. 1979. Vol. 21. №2. P. 152-153. <http://dx.doi.org/10.1007/bf02909468>